

$$\frac{1}{T_{1d \text{ intra}}} = 8.4 \times 10^{-3} \text{ s}^{-1}; \frac{1}{T_{1d \text{ inter}}} = 2.4 \times 10^{-3} \text{ s}^{-1} \quad (3)$$

The total dipolar contribution thus amounts to about 92.5 s at room temperature.

According to Hubbard³ the spin-rotation contribution is given by

$$\frac{1}{T_{1sr}} = \frac{8\pi^2 I_0 k T}{3 \hbar^2} (2c_{\perp}^2 + c_{\parallel}^2) \tau_{sr} \quad (4)$$

where the correlation time for the molecular angular velocity, τ_{sr} , is related to τ_d by the expression $\tau_d \tau_{sr} = I_0 / 6kT$. These equations are valid for a spherical molecule undergoing isotropic Brownian reorientation when $\tau_d \gg \tau_{sr}$. With $I_0 = 7.88 \times 10^{-38} \text{ cm}^2 \text{ g}$ which is the principal moment of inertia for P_4O_6 , we get $\tau_{sr} = 3.8 \times 10^{-16} \text{ s}$.

Assuming that the spin-lattice relaxation is controlled by the dipolar and spin-rotation interactions we obtain $T_{1sr} = 20.8 \text{ s}$ since $T_1 = 17 \text{ s}$ and $T_{1d} = 92.5 \text{ s}$ at 21°C. Eqn. (4) then gives 13.0 kHz for the spin-rotation interaction constant which is a rather small value.⁴ It is often found that the Debye formula gives a maximum value for τ_d . In some cases values for τ_d which are from two to six times smaller than those obtained by the Debye expression have to be postulated.^{4,5} If that is the case for P_4O_6 the spin-rotation interaction will be the dominant relaxing mechanism and thus yielding 14.4 kHz for the spin-rotation interaction constant. A few spin-lattice relaxation time measurements on liquid P_4 have been reported⁶ and T_1 was found to fall with temperature. This was not explained satisfactorily; however, the possibility of a spin-rotation interaction was mentioned. No definite conclusion concerning P_4O_6 can be drawn, however, before T_1 measurements have been performed over an extended temperature range.

2. *The solid state.* The ^{31}P spin-lattice relaxation time of solid phosphorus trioxide has been measured at 9 MHz in the temperature range 21°C to -19°C (see Fig. 1). T_1 increases very rapidly from 40 s to 290 s for temperatures down to 14°C. Thereafter T_1 levels off gradually and reaches a value of about 600 s at -19°C. The steep rise in T_1 just below the melting point indicates that the first two T_1 values actually were measured in the premelt. The line width was broad and seemed to change only slightly in the temperature range studied. This indicates that the translational diffusion can be neglected and that the relaxa-

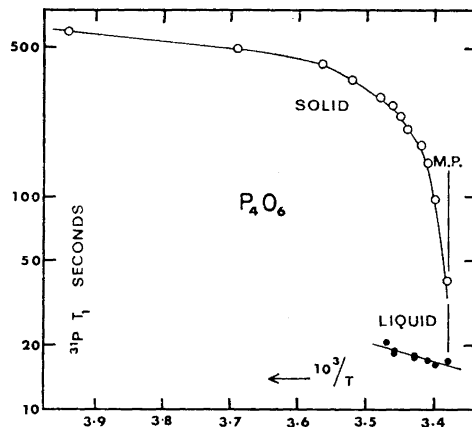


Fig. 1. The ^{31}P spin-lattice relaxation time in supercooled and solid P_4O_6 at 9 MHz. M.P. stands for melting point.

tion probably is caused by interactions modulated by molecular reorientation.

It is known that simple tetrahedral molecules exhibit nearly isotropic reorientation in the solid state.⁷ This is also found to be the case for the P_4 tetrahedra.^{8,7} The P_4O_6 molecules are reasonably spherical with a 'radius' of about 3.2 Å. The assumption of a fairly unhindered rotation of the molecules in the solid at temperatures not too far from the melting point, seems therefore plausible.

If an isotropic Brownian reorientation is assumed in the solid, the dipolar contribution to the spin lattice relaxation is easily found to be²

$$\frac{1}{T_1} = \frac{9\gamma_p^4 \hbar^2}{20\pi^2 r_{p-p}^6 \omega_p^2 \tau_d} = 1.3 \times 10^{-8} \tau_d^{-1} \quad (5)$$

at 9 MHz since $\omega_p \tau_d \gg 1$ in the temperature region considered. This gives the correct temperature dependence for τ_d , and the graph of $\ln \tau_d$ against $1/T$ could then consist of two straight lines of different activation energies. This sounds reasonable if the isotropic rotational motion of the molecules 'freezes out' gradually and becomes more restricted around 14°C. Using eqn. (5) τ_d is found to be of the order of 10^{-5} to 10^{-6} s in the measured temperature range. It would be interesting to measure T_1 at a different frequency to see if T_1 is frequency dependent as eqn. (5) predicts.

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Determination of the Rate Constants for Alkylation of DNA *in vitro* with Methanesulfonic Esters

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The mutagenic alkylating agents, methyl, ethyl, and isopropyl methanesulfonate (MMS, EMS, and iPMS, respectively) give quantitatively and qualitatively different effects both with biological materials^{1,2} and with DNA *in vitro*.^{3,4} The site of alkylation of DNA for these substances is supposed to be at different groups, *i.e.* unesterified phosphate oxygens and purine base nitrogens.^{5,6}

As a basis for a quantitative comparison of the effects of treatment with different alkylating agents, the reaction rate constants with DNA *in vitro* were determined. The site of alkylation will be discussed.

Experimental. Calf thymus DNA was purchased from Sigma Chemical Co. Tritium labelled MMS, EMS and iPMS were synthesized according to Wachtmeister *et al.*⁷ Unlabelled

MMS and EMS from Eastman Organic Chemicals and iPMS from Koch-Light Laboratories were used.

The labelled alkylating agents were diluted with inactive substance to the following specific activities: 8 mCi/mole for MMS, 50 mCi/mole for EMS, and 70 mCi/mole for iPMS. The concentration of MMS and EMS were 3 mM and for iPMS 0.5, 0.75, and 1.5 mM. DNA was alkylated at a concentration of 1 mg/ml in a 0.02 M phosphate buffer of pH 7.0 at 25°C. Samples of 1 ml were withdrawn at different times. iPMS, which has a half life of 1 h at 25°C,¹ was incubated for 8 h with DNA, *i.e.* to practically complete consumption of the ester. DNA was precipitated by 2 volumes of 95 % ethanol and washed four times with 70 % ethanol. It was then redissolved and hydrolyzed in 1 N HCl for 20 min at 100°C in vacuum. The samples were mixed with a scintillation solution consisting of dioxane-naphthalene-PPO-POPOP⁸ to which was added 0.5 ml 1 N HCl and 0.5 ml of hydroxide of hyamine 10X (Packard Co.). The liquid scintillation spectrometer used was a Nuclear Chicago Unilux model 6851 (Packard Co.).

Results. During incubation with DNA in a buffer solution an alkylating agent reacting according to S_N2 (in the present case MMS and EMS^{1,5}) is used up mainly in additive reactions with the nucleophiles present. Besides water and DNA, the phosphate ions are alkylated. Thus, the alkylating agent disappears with the rate constant k'

$$k' = [\text{H}_2\text{O}] \cdot k_{\text{H}_2\text{O}} + [\text{HPO}_4^{2-}] \cdot k_{\text{HPO}_4^{2-}} + [\text{H}_2\text{PO}_4^-] \cdot k_{\text{H}_2\text{PO}_4^-} + [\text{DNA-P}] \cdot k_{\text{DNA}}$$

where [DNA-P] is the concentration of DNA in mole/l nucleotide phosphorus. Under the reaction conditions used the factors $[\text{H}_2\text{PO}_4^-] \cdot k_{\text{H}_2\text{PO}_4^-}$ and $[\text{DNA-P}] \cdot k_{\text{DNA}}$ are negligible. $k_{\text{H}_2\text{PO}_4^-}$ is *ca.* 17 times less than $k_{\text{HPO}_4^{2-}}$ and [DNA-P] is about ten times lower than $[\text{HPO}_4^{2-}]$.⁹ The constants for secondary phosphate and water have been determined by Osterman-Golkar *et al.*¹⁰

iPMS reacts predominantly according to an S_N1 mechanism, especially with weak nucleophiles,^{5,8,10} and its total decay rate is therefore uninfluenced by nucleophiles present, although it may be affected by salts.⁹ In the absence of competing reactions and at low concentrations of a dissolved nucleophile, *i.e.* under conditions which apply to DNA in the present case, it is, however, possible to assign formally